

REMARKS

The Examiner is thanked for conducting a telephone interview on July 6, 2011 with the undersigned. It is believed that the interview helped to further the prosecution by identifying the outstanding issues and ways to resolve them.

Claims 2-5, 12, 13 and 18-21 are pending.

At page 2 of the Action, claims 2-5, 18 and 19 are rejected under 35 U.S.C. § 102(b) as being anticipated by Takayanagi (US 5,945,564).

At page 2 of the Action, claims 2-5, 19 and 21 are rejected under 35 U.S.C. § 102(b) as being anticipated by Yoshida et al (US 5,489,572).

At page 2 of the Action, claims 2-5, 19 and 21 are rejected under 35 U.S.C. § 102(b) as being anticipated by Kuramochi et al (US 5,661,111).

At page 3 of the Action, claims 2-5, 19 and 21 are rejected under 35 U.S.C. § 102(b) as being anticipated by Tanaka et al (US 5,298,482).

At page 3 of the Action, claims 12 and 13 are rejected under 35 U.S.C. § 103(a) as being unpatentable over Daisuke et al (JP 5310657), taken with Takayanagi, Kuramochi et al, and Tanaka et al.

As discussed in the interview, Applicants submit that the above five rejections should be withdrawn because the cited references do not enable making the presently claimed ALA phosphate salt and, thus do not disclose or render obvious the present invention, either alone or in combination.

In the interview, Applicants explained that none of Takayanagi, Yoshida et al, Kuramochi et al and Tanaka et al enables making the presently claimed phosphate compounds, and therefore cannot stand against the novelty of the present claims.

As already discussed in the Amendment under 37 C.F.R. § 1.111 filed June 23, 2010, Takayanagi, while mentioning phosphates of (dideutero)ALA at col. 2, line 61, does not disclose any workable process for actually manufacturing the specific ALA phosphates of formula (I) of present claim 2.

As one allegedly suitable reaction, the acidic hydrolysis of compound (C) shown in the reaction scheme at col. 3 of Takayanagi is disclosed at col. 4, lines 26-32. However, as can be inferred from the disclosure therein, the acid to be used in this acidic hydrolysis must be a strong acid, such as HCl, HBr or a strong organic acid such as methane- or p-toluene sulfonic acid. Thus, it is clear to one skilled in the art that this acidic hydrolysis cannot be performed with a weak acid such as phosphoric acid to obtain the presently claimed phosphate salt.

A second method for making the compound (I) of Takayanagi is shown in the reaction scheme at cols. 4-5. However, it can be seen that for making compound (I) from compound (H) in this reaction very vigorous reaction conditions are required, and it would have been clear to one skilled in the art that phosphoric acid would not be a suitable reactant in this reaction (col. 6, lines 46-57). Also, it is noted that from this reaction only compounds are obtained where the ALA derivative does not have a free carboxylic acid group, but rather a carboxyl group esterified by a group R¹. Therefore, this process does not provide a suitable or workable way to obtain the presently claimed phosphate salt.

In view of the above, Takayanagi does not provide an enabling disclosure for making the presently claimed phosphate compounds, and therefore cannot stand against the novelty of the present claims.

Yoshida et al, Kuramochi et al and Tanaka et al mention ALA phosphate but do not teach a process for making ALA phosphates. Furthermore, no process is disclosed in these

references according to which the presently claimed phosphate compounds could be obtained. Thus, these references do not provide an enabling disclosure for making the presently claimed phosphate compounds.

In the interview, Applicants also explained that ALA is unstable, as shown by the article, Novo, et al, "Chemical instability of 5-aminolevulinic acid used in the fluorescence diagnosis of bladder tumors" (Journal of Photochemistry and Photobiology B: Biology 34 (1996) 143-148), and as a result, manufacture of the ALA phosphate salt would not have been routine in the art.¹ Furthermore, in response to the Examiner's question as to whether Novo et al teaches making the claimed ALA phosphate salt, albeit unpurified, Applicants confirm that Novo et al does not produce the claimed ALA phosphate. Novo et al describes a method of preparing ALA aqueous solutions, but does not teach that ALA phosphates are made. At page 144, 1st column, 1st paragraph under "Experimental Results," Novo et al discloses "All solutions were buffered using a ten-times concentrated Dulbecco's phosphate-buffered saline (Gibco BRL) to obtain the phosphate and salts concentrations usually used for biological media (total concentration of phosphates, 9.6 mM; ...)." Dulbecco's phosphate-buffered saline is a phosphate buffer and contains a phosphoric acid and salt. However, it is apparent that the salts do not represent an ALA salt or an ALA phosphate, but represent the salts of Na, K and the like that constitute Dulbecco's phosphate-buffered saline, <http://www.dspbio.co.jp/pdf/product/18-604-54.pdf>.

The article describes a method of preparing ALA aqueous solutions of pH 5.19, pH 6.31 and pH 7.44.

¹ A copy of Novo et al, inadvertently not submitted previously is being submitted herewith, and the Examiner is thanked for agreeing to consider Novo et al. after final rejection.

Specifically, the ALA hydrochloride (HCl) is dissolved in tenfold concentrated Dulbecco's phosphate-buffered saline, and the pH is adjusted with NaOH or HCl. However, there is no disclosure on how such components are added in the preparation of the ALA aqueous solutions.

Based on the fact that the pH can be adjusted with HCl, it is highly probable that HCl is used in the preparation of the ALA aqueous solution having a pH of 5.19, which is the solution having the lowest pH among the three ALA solutions disclosed.

As seen from Fig. 3 of Novo et al, ALA is unstable at pH 6.13 or more, and becomes 2,5-(β -carboxyethyl)dihydropyrazine (CHPY), then 2,5-(β -carboxyethyl)pyrazine (CRY).

The only stable solution in Novo et al is the ALA aqueous solution of pH 5.19, which contains HCl. The other ALA aqueous solutions also contain HCl as a counter of ALA in ALA hydrochloride. The point is that the ALA aqueous solution of pH 5.19 contains a large amount of HCl, compared with the molar amount of ALA.

The details of the amounts of each component in the ALA aqueous solution of pH 5.19 are unclear, but the ALA aqueous solution of pH 5.19 contains, other than ALA, a large amount of cations, such as Na or K, and a large amount of anions, such as Cl, H₂PO₄ or HPO₄.

However, because the solution is an aqueous solution, there is a higher possibility that each ion is free as opposed to forming a salt. Further, even if a salt forms, the salt most probably is the salt of HCl, which has the highest acidity and is present in an equivalent amount or more to the molar amount of ALA. Also, although there is a lower possibility, the next candidate for salt formation is H₂PO₄. However, this possibility is extremely low.

Since ALA decomposes at pH 6.31 or more, the manufacture thereof is extremely difficult.

In contrast, according to the present invention, ALA can be obtained by purification under specific conditions with ion exchange resin, whereby ALA phosphate is manufactured. Nonetheless, should the Examiner have a different view of the teachings of Novo et al., Applicants will amend the claims as the Examiner suggested might be helpful, to recite that the present salt is “isolated and purified.”

In the interview, Applicants further discussed the evidence that shows that 5-aminolevulinic acid phosphate alone was not isolated and purified prior to March 28, 2005, the international filing date of the present application; and that 5-aminolevulinic acid phosphate alone was isolated and purified for the first time by the present inventors. Specifically, Applicants have searched “pentanoic acid, 5-amino-4-oxo-, phosphate (1:1) (5-aminolevulinic acid phosphate)” in the Data Base of CAS (Chemical Abstracts Service: <http://www.cas.org/>), and as a result, Applicants were able to confirm that the 5-aminolevulinic acid phosphate (ALA phosphate) was, for the first time, registered as No. 868074-65-1 on November 15, 2005.²

Still further, in the interview Applicants explained that the claimed compounds have unexpectedly superior properties. Specifically, the examples of the present specification, particularly Examples 3-6 and Comparative Examples 1-3 at pages 18-21, establish that the presently claimed ALA phosphates exhibit a reduced smell and improved organoleptic properties as compared with the ALA•HCl known in the art, even when the ALA•HCl was produced according to the same procedure as disclosed in the present specification (using hydrochloric acid instead of the phosphoric acid of formula (III)). Applicants maintain that this is a significant improvement in product quality, especially in the field of medical compositions,

² A copy of the search results inadvertently not submitted previously is being submitted herewith, and the Examiner is thanked for agreeing to review the results after final rejection.

where a patient can be repulsed by an unpleasant smell and/or taste of a medical composition; and this improvement, achieved by the present invention, could not have been foreseen by one skilled in the art based on the teachings of the cited prior art. Thus, the claimed ALA phosphate has unexpectedly superior properties as compared to the known ALA•HCl.

In summary, the cited references do not enable making the claimed ALA phosphate. This is further supported by Novo et al who teaches that ALA is unstable and by Applicants' search results for ALA phosphate. Additionally, the presently claimed ALA phosphate compounds have unexpectedly superior properties as compared to the known ALA•HCl. Accordingly, reconsideration and withdrawal of all the §§102(b)/103(a) rejections based on the cited references are respectfully requested.

At page 4 of the Action, claim 20 is objected to as being dependent upon a rejected base claim, but is indicated to be allowable if rewritten in independent form.

Claim 20 is patentable in its present form because claim 2, from which claim 20 depends, is patentable as discussed above.

Allowance is respectfully requested. If any points remain in issue such as a belief that the claims should be amended to recite that the ALA phosphate salt is "isolated and purified," the Examiner is kindly requested to contact the undersigned at the telephone number listed below.

The USPTO is directed and authorized to charge all required fees, except for the Issue Fee and the Publication Fee, to Deposit Account No. 19-4880. Please also credit any overpayments to said Deposit Account.

Respectfully submitted,

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Chemical instability of 5-aminolevulinic acid used in the fluorescence diagnosis of bladder tumours

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Abstract

Aqueous solutions of 5-aminolevulinic acid (ALA) prepared for intravesical instillation in the framework of a clinical study on the fluorescence diagnosis of urothelial bladder cancer were found to be unstable. This chemical instability of ALA was studied in aqueous solution of 37 °C as a function of coprecipitation, pH and reaction time. Our investigations showed that the reaction of ALA is an irreversible process, which yields at least two reaction products in the pH range studied (pH lower than 9): the 2,5-(β -carboxyethyl) dihydropyrazine and the 2,5-(β -carboxyethyl) pyrazine. As a result of these studies, the conditions for the preparation of ALA solutions to be used for intravesical instillation were optimized: solutions of ALA in phosphate-buffered saline at a concentration of 0.18 M (3 g of ALA in 100 ml) neutralized to pH 5, were prepared and stored on ice until use. Solutions prepared under these conditions were stable and were used for fluorescence diagnosis of bladder tumours with successful results. The effect of the pH and the composition of the urine on the extent of the reaction of ALA and on the nature of its reaction products formed during instillation was investigated by comparing the urine of patients before and immediately after instillation of ALA.

Keywords: Chemical instability; 5-Aminolevulinic acid; Fluorescence diagnosis; Bladder tumours; Cancer

1. Introduction

5-aminolevulinic acid (ALA) has been found to induce the formation of photosensitizing concentrations of endogenous protoporphyrin IX (PPIX) in certain types of cell and tissue [1,2]. Because of advantages such as tissue specificity, rapid clearance of induced photosensitizer and possibility of topical administration of ALA-induced PPIX compared with exogenous photosensitizers, ALA has been extensively used in the last years in both pre-clinical and clinical studies on photodynamic therapy (PDT) and fluorescence diagnosis.

Our group is involved in different clinical studies on photodynamic therapy and fluorescence diagnosis with ALA-induced PPIX, conducted in collaboration with several departments of the Medical University in Lübeck [3,4]. In all these clinical studies, ALA is applied to the tissue at high concentrations and is usually neutralized to a pH well tolerated by the tissue [5]. During the preparation of aqueous solutions of ALA under the conditions used for instillation into the human bladder, the solutions turned from colourless to yellow within minutes as they were buffered to neutral pH. Furthermore, the colour of the solutions became more intense with time and their pH decreased significantly even in the presence of a phosphate buffer. These changes revealed the

occurrence of a chemical reaction of ALA which is not desirable for clinical application owing to the practical problems derived and, more important, owing to the possible decrease in the efficacy of the instillation. Experiments were therefore performed to study the reaction of ALA in aqueous solution as a function of different parameters (pH, concentration, temperature and time) in order to determine the conditions under which ALA is stable, to find out whether the reaction is reversible and to characterize the reaction products.

It is well known that *in vivo* ALA undergoes an enzymatically induced dimerization to give porphobilinogen in the biosynthetic pathway to haem [6]. However, in the absence of the corresponding enzymes, the spontaneous dimerization of ALA yields different cyclic products. Granick and Manczak [7] reported the dimerization of ALA to 2,5-(β -carboxyethyl) dihydropyrazine (CHPY) in alkaline solution. Later, Franck and Stratmann [8] showed that in alkaline solution the condensation of two molecules of ALA leads to two products in the ratio of 1:10, the minor product being identified as porphobilinogen and the predominant product as the unstable and difficult-to-isolate CHPY. Recently, Butler and George [9] identified three condensation products of ALA in aqueous solution depending on the reaction condi-

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tions: the CHPY, which is formed at moderate pH under anaerobic conditions and yields 2,5-(β -carboxyethyl)-pyrazine (CPY) by aerial oxidation, and pseudo-porphobilinogen, obtained in strongly alkaline conditions together with the diflavydopyrazine. The data of these workers indicated that no porphobilinogen is formed in the non-enzymatic cyclization of ALA.

These previous studies showed that ALA reacts in aqueous solution to yield several condensation products. However, the controversial results obtained by the different workers show the complexity of the reaction of ALA, typical of carbonyl group reactions [10]. The conditions (concentration, temperature, etc.) necessary for this reaction to take place were not investigated, so that the stability range of ALA is not yet defined. Furthermore, it must be proved that the products of the reaction of ALA after long reaction times (6 h) at high temperatures (70–100°C) are the same as those which form in the presence of urine and at much milder conditions, e.g. during the instillation into the bladder of patients (1–2 h at 37°C). Therefore the reaction of ALA in the present study has been investigated in aqueous solution under the latter conditions in order to identify the reaction products and to estimate the stability of ALA as a function of pH and concentration during the times relevant for instillation. This study was performed using the UV-visible absorption spectroscopy technique.

2. Experimental details

Aqueous solutions of ALA at different pH and concentration values were prepared by dissolving the appropriate amount of 5-aminolevulinic acid hydrochloride (Merck) in bidistilled water and adding NaOH (Merck, p.a.) to obtain the required pH. All solutions were buffered using a ten-times concentrated Dulbecco's phosphate-buffered saline (Gibco BRL) to obtain the phosphate and salts concentrations usually used for biological media (total concentration of phosphates, 9.6 mM; concentration of additional salts, 0.140 M). Saline without bivalent cations was used to avoid the formation of ALA precipitates in basic media. HCl (Merck) was added if necessary to acidify the solutions of ALA. Anaerobic solutions were prepared by bubbling nitrogen into the aqueous solution for 1 h before adding ALA.

A clinical study on the fluorescence diagnosis of bladder tumours [3] was performed on ten patients using solutions of ALA prepared under optimized conditions derived from the studies in aqueous solution. For the first two patients, solutions of ALA at pH 5.5 were used. The good results obtained in these cases, i.e. no irritation or damage of the bladder tissue and a pronounced fluorescence of ALA-induced PPIX, led to a further decrease in the pH to pH 5.0. The concentration of the ALA solutions was 0.18 M in all cases and a volume of 50 ml was instilled. The instillation time was 1 h for the first two patients and 2 h for the other eight patients. Samples of urine were collected before and

immediately after the instillation of ALA into the bladder of the patients and were kept on ice until their analyses. Note that the samples of urine collected after the instillation of ALA (ALA-urine) contained, in addition to the urine produced during the instillation, the ALA which was not taken up by the bladder cells and the products of the non-enzymatic condensation of ALA formed during the instillation. Dilute solutions (1/100) of the urine collected after the instillation of ALA were prepared immediately after collection and were also kept on ice. The urine collected before the instillation was diluted 200 times to allow the measurement of the absorption spectra.

pH was measured with a WTW pH meter (model 521) equipped with a combined pH electrode (WTW type E56). Absorption spectra were measured with a UV-visible spectrometer Lambda 14P from Perkin-Elmer.

3. Results and discussion

3.1. Reaction of 5-aminolevulinic acid in aqueous solution

Stock solutions of ALA at pH 7.3–7.4 and different concentrations (0.005 M, 0.18 M and 0.30 M) were prepared and kept at 37°C. The two more concentrated solutions turned yellow when being neutralized and their pH decreased rapidly. 3 min after preparation, aliquots of the stock solutions were taken to obtain 5 mM dilute solutions of pH 7.3, whose absorption spectra are shown in Fig. 1(a). The spectrum of the 5 mM stock (full curve) is identical with the absorption spectrum of ALA, whereas the spectra corresponding to the 0.18 M and 0.30 M stock solutions show a new band at about 350 nm, as well as changes in the form of the spectrum at shorter wavelengths. These changes in colour, pH and absorption spectra of the concentrate stocks reflect the occurrence of a chemical reaction of ALA. Furthermore, this reaction is largely dependent on the initial concentration of ALA, but it cannot be attributed to a simple aggregation process, since the spectral changes are permanent after dilution of the concentrated stocks.

Using the same dilution procedure, the absorption spectra of the three samples were measured 1 h after preparation of the stocks (Fig. 1(b)). After this time interval, the spectrum of the 5 mM stock solution already differs from that of ALA, showing analogous changes to the spectra of the concentrated solutions after reaction for 3 min, which can be explained as above. However, a new band at about 275 nm becomes distinct in the spectra corresponding to the concentrated stocks after reaction for 1 h. For a given initial concentration of ALA, the ratio of the absorbance at 275 nm to that at 350 nm increases with increasing reaction time, although the decrease in the concentration of ALA due to the reaction causes a decrease in the absorbance at 275 nm and has no effect on the absorbance at the 350 nm band. These facts can only be interpreted by the contribution of at least two different absorbing species to the experimental absorption spectra, so

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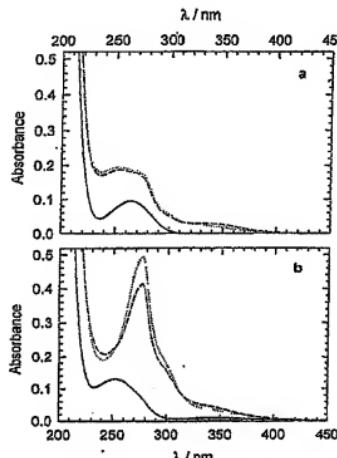


Fig. 1. Absorption spectra of ALA solutions (5 mM; pH 7.3) which were prepared from ALA stocks of pH 7.3–7.4 at three different concentrations, namely 5 mM (—), 0.18 M (—) and 0.30 M (· · ·), after reaction (a) for 3 min at 37 °C and (b) for 1 h at 37 °C.

that the absorption band at 275 nm must be due to a second product of the reaction of ALA. Studying the effect of oxygen on the absorption spectra of a reacting solution of ALA allowed us to assign these two absorption bands to the reaction products identified by Budde and George [3]. The absorption spectrum of a solution of ALA reacting under anaerobic conditions (Fig. 2) shows an intense band at about 350 nm and a shoulder at the low-wavelength absorption band, which must be attributed to the dihydropyrazine CHPY, the only product of ALA reaction in the absence of oxygen. When a small amount of oxygen was allowed to enter into the stock solution, the absorption spectrum changed significantly (Fig. 2), decreasing the absorbance at the 350 nm band as a new band appears at about 275 nm, as observed for the aerated solutions. These facts are perfectly explicable by the oxidation of CHPY to give CPY, so that the concentration of CHPY decreases as CPY is formed, this product being responsible for the absorption band at 275 nm which increases in intensity as the concentration of CPY becomes higher.

In addition to the concentration dependence, the occurrence of the reaction of ALA was found to be strongly dependent on the pH. When ALA is dissolved in water without

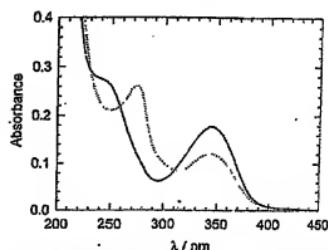


Fig. 2. Absorption spectra of ALA solutions (4.6 mM; pH 7.3), which were prepared from a deaerated stock of pH 7.9 and concentration 0.27 mM reacting at room temperature: —, after reaction for 5 min; · · ·, after letting some oxygen enter into the stock.

adding NaOH, the acid solution obtained (0.3 M; pH 2) is stable, showing an absorption spectrum which is invariant with time and has no band at wavelengths longer than 320 nm. The reaction of ALA in the pH range relevant for clinical use was studied by preparing stock solutions of ALA of the same concentration (0.18 M) and different pH values (5.2, 6.3 and 7.4), which were left to react at 37 °C for several hours. It was observed that, although the same concentration of phosphate buffer was present in all three solutions, the acidity of the stock at pH 5.2 did not vary during the reaction, whereas the pH values of the other two stocks decreased significantly, as indicated in Fig. 3(a). Since the absorbances of the stock solutions both at 350 nm and at 275 nm were too high to be measured, the absorbance of the CHPY was detected at 400 nm instead of 350 nm as a function of the reaction time (Fig. 3(a)). Dilute solutions (5 mM; pH 7.3) were thus prepared from each stock at different reaction times in order to measure the absorbance at 275 nm (Fig. 3(b)). Since neither ALA nor CPY absorbs at 400 nm, the absorbance measured at this wavelength is proportional to the concentration of CHPY formed in the reaction of ALA. The data in Fig. 3(a) show that the concentration of CHPY increases very rapidly in the first 10–15 min of reaction at any pH, indicating that the rate of formation of this product is high. However, about ten times more product is formed during that reaction time in the pH 7.4 stock solution than at pH 5.2, and two times more than in the stock of pH 6.3. This suggests that the rate of formation of CHPY is strongly dependent on the pH and this may be due to the different reactivities of the protonated and deprotonated forms of ALA and/or to an acid-base catalysis of the reaction. At longer reaction times the behaviour of the 400 nm absorbance is different for the three stock solutions. It increases slowly with increasing reaction time in the solution at pH 5.2, and in contrast decreases slightly in the pH 6.3 stock and significantly in the pH 7.4 stock. These different variations in the concentration of

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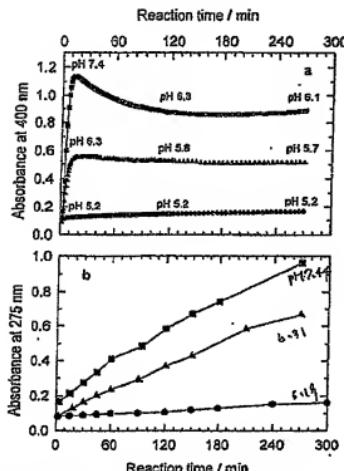


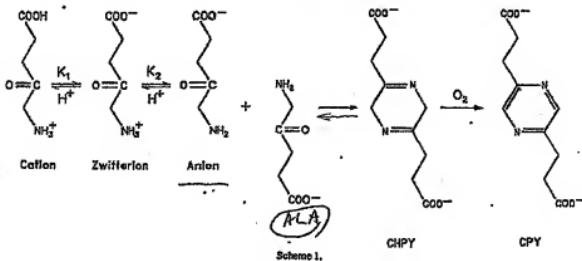
Fig. 3. Time dependence of the reaction of ALA at 37 °C for solutions of concentration 0.18 M and different initial pH values. (a) Absorbances at 400 nm versus reaction time of ALA at pH 7.4 (●), pH 6.3 (○), pH 6.1 (△), pH 5.8 (×), pH 5.7 (×), pH 5.2 (□), and pH 5.2 (■). The pH of the stock solutions is indicated in three random times (65 min). Absorbances at 275 nm measured for the dilute solutions (5 mM; pH 7.3) which were prepared from those stocks at different reaction times.

CHPY as a function of reaction time can be explained by the further reaction of CHPY to give CPY, whose rate would

depend on the concentration of CHPY and therefore would be higher as the pH of the stock increases. The data of the 275 nm absorbances (Fig. 3(b)) are in keeping with this interpretation. The rate of increase in the absorbance measured as the slope of the curves is ten and 20 times higher for the solutions of pH 6.3 and pH 7.4 respectively, than for the pH 5.2 stock. This indicates that the rate of formation of CPY increases strongly with increasing pH.

The potential reversibility of the reaction of ALA was checked by comparing the absorption spectra of two acid diluted solutions of ALA: one prepared from an acid and stable stock solution and the other from a reacting stock (data not shown). These spectra were very different: the former coincided with the absorption spectrum of ALA at low pH, whereas the latter showed the presence of the reaction products of ALA. Hence, the reaction of ALA is not reversible by acidification.

The pH dependence of the reaction of ALA can be explained on the basis of the acid-base equilibria of this aminoacid (Scheme 1). The values of the corresponding acidity constants were determined spectrophotometrically and are $pK_1 = 4.05 \pm 0.05$ and $pK_2 = 8.3 \pm 0.1$. These values indicate that the zwitterion is the major species present in the pH range between 3 and 7.5, although significant amounts of the two other acid-base species exist depending on the acidity. Thus, at pH 5, about 10% of the molecules of ALA are cations whereas, at pH 7.3, about 10% of the ALA molecules are anions. According to these results, a scheme similar to that proposed by Butler and George [9] could explain the reaction of ALA, where the anion, a species with a deprotonated amino group, is the only one that is able to react with the ketone group of a neighbouring molecule to yield the cyclic dihydropyrazine (Scheme 1). For the condensation to occur, the amino group of ALA should be deprotonated. This explains the strong pH dependence of the reaction, since the concentration of the anion increases with the pH, as shown above. Therefore the solutions of ALA are only stable at such high acidities where the anion does not exist (note that,



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although at pH 5.2 the concentration of that species is only 0.07% of the total concentration of ALA, some reaction does occur as shown in Fig. 3). Furthermore, as the anion reacts, the concentration of protons increases in the solutions of ALA in order to keep the equilibrium zwitterion-anion (see Scheme 1), explaining the decrease in the pH observed during the reaction of ALA. Independent of the mechanism of the reaction, the study of which is beyond the objectives of this work, it can be expected that the condensation of two molecules will be strongly influenced by the concentration, temperature and viscosity. Those parameters together with the acidity must be optimized and controlled in order to avoid the reaction of ALA used in clinical studies of PDT and fluorescence diagnosis.

3.2. Reaction of 5-aminolevulinic acid during instillation in the human bladder

On the basis of our results, conditions for the preparation of ALA solutions to be instilled into the bladder of patients for the fluorescence diagnosis of bladder tumours were optimized. Since the concentrations of ALA for clinical use should be high enough to induce the accumulation of detectable amounts of PPIX, a minimal concentration of 0.18 M (3 g of ALA in 100 ml of phosphate buffer) was used. The pH of the solutions was reduced to pH 5, which is near to the limit of tolerance of the bladder, since the pH range of human urine can vary from 4.8 to 8.4 [11]. Finally, the temperature should be controlled by cooling the stocks used for the preparation of the solution of ALA, preparing the mixture on ice and storing the resulting solution at low temperatures. Solutions of ALA prepared and kept under these conditions were stable for several days.

In order to study the reaction of ALA during instillation in the human bladder, absorption spectra and pH of the patients' urine collected before and immediately after the instillation were measured. The pH of urine collected before the instillation differed significantly among the different patients, varying from pH 5.1 to pH 7.7. A general decrease in pH was observed in the urine after the instillation of ALA. However, for those patients whose urine showed a pH higher than 6.5 before instillation, the pH of the ALA-urine was still higher than 6. This means that the urine produced during the instillation can significantly increase the pH of the solution of ALA, thus leading to a higher rate of non-enzymatic condensation of ALA.

Difference spectra were obtained by subtraction of the absorption spectra of urine collected before the instillation of ALA from those of the ALA-urine, previously normalized at short wavelengths. Fig. 4(a) shows a typical difference spectrum (dotted curve) compared with the absorption spectrum of a dilute solution obtained after reaction for 2 h at 37 °C of a 3% stock of ALA of pH 6.3 (full curve), which accounts for the reaction in aqueous solution under similar conditions (these two spectra have been normalized at 277 nm to allow comparison). The similarities between the dif-

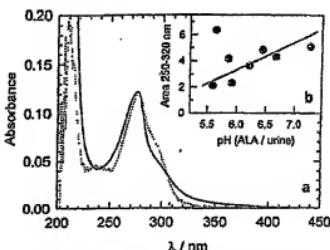


Fig. 4. (a) Absorption spectrum of a dilute aqueous solution of ALA (5 mM; pH 7.3) prepared from a stock of concentration 0.18 M and pH 6.3 (solid line), and difference spectrum obtained from the absorption spectra of the urine collected before the instillation of ALA from that of the ALA-urine (dotted line). The spectra were normalized at 277 nm to allow comparison. (b) Plot of areas between 250 and 320 nm of the difference spectra obtained as explained above vs. pH of the ALA-urine for the patients with 2 h of instillation.

ference spectra and that obtained in aqueous solution suggest that the pyrazine CPY is also formed by reaction of ALA in urine. The slight differences might be attributed to an effect of the medium or to further reaction of the CPY with constituents of the urine. The accuracy of the difference spectra is not high enough to detect the absorption of the CPY.

The areas between 250 nm and 320 nm of those difference spectra were determined and a correlation with the pH of the ALA-urine was found (Fig. 4(b)). The absorption of the reaction product increased with increasing pH. This result shows again the necessity to control the pH not only before but also during the instillation in order to avoid the non-enzymatic reaction of ALA. These variations could be avoided by adding a sufficient amount of a suitable buffer to the solution of ALA, or by controlling the pH of the urine of the patients by means of a suitable diet.

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Our special thanks go to Drs. M. Mosquera and F. Rodríguez Prieto from the University of Santiago de Compostela (Spain) for their critical review of the manuscript. M.N. thanks the Xunta de Galicia for a post-doctorate scholarship.

References

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 L1 1 868074-65-1/RN

=> d

L1 ANSWER 1 OF 1 REGISTRY COPYRIGHT 2009 ACS on STN
 RN 868074-65-1 REGISTRY
 ED Entered STN: 15 Nov 2005
 CN Pentanoic acid, 5-amino-4-oxo-, phosphate (1:1) (CA INDEX NAME)
 OTHER NAMES:
 CN 5 -Aminolevulinic acid phosphate
 CN 5-Aminolevulinic acid phosphate
 MF C5 H9 N O3 . H3 O4 P
 SR CA
 LC STN Files: CA, CAPLUS, CASREACT, TOXCENTER, USPATFULL

CM 1

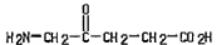
CRN 7664-38-2
 CMF H3 O4 P



2/5

STN Tokyo

CM 2

CRN 106-60-5
CNF C5 H9 N O34 REFERENCES IN FILE CA (1907 TO DATE)
4 REFERENCES IN FILE CAPLUS (1907 TO DATE)

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USPTO MANUAL OF CLASSIFICATIONS THESAURUS ISSUE DATE: Aug 2009

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L2 4 L1
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L2 ANSWER 1 OF 4 CA COPYRIGHT 2009 ACS on STN
[Full Text](#)

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STN Tokyo

AN 148:269318 CA
 TI Novel crystal of 5-aminolevulinic acid phosphate and process for
 production thereof

TIJP 5-アミノレブリン酸リン酸塩の新規結晶、その生成のためのプロセス

[機械翻訳]

IN Tachiya, Naohisa
 PA Cosmo Oil Co., Ltd., Japan
 SO PCT Int. Appl., 27 PP.
 CODEN: PIKXD2
 DT Patent
 LA Japanese
 FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI WO 2008020532	A1	20080221	WO 2007-JP64515	20070724
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JP 2008044882	A	20080228	JP 2006-221538	20060815
AU 2007285257	A1	20080221	AU 2007-285257	20070724
CA 2650206	A1	20080221	CA 2007-2650206	20070724
EP 2053039	A1	20090429	EP 2007-791240	20070724
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IN 2008DN09356	A	20090320	IN 2008-DN9356	20081107
KR 2009042757	A	20090430	KR 2008-728355	20081120
MX 2008015159	A	20081212	MX 2008-15159	20081127
NO 2008005283	A	20090313	NO 2008-5283	20081217
CN 101472879	A	20090701	CN 2007-80022901	20081219
PRAI JP 2006-221538	A	20060815		
WO 2007-JP64515	W	20070724		

OS MARPAT 148:269318

RE.CNT 17 THERE ARE 17 CITED REFERENCES AVAILABLE FOR THIS RECORD
 ALL CITATIONS AVAILABLE IN THE RE FORMAT

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L2 ANSWER 2 OF 4 CA COPYRIGHT 2009 ACS on STN

Full Text

AN 147:386245 CA

TI Method for manufacturing phosphate salt of amino acid or its ester

TIJP アミノ酸リン酸塩の製造方法 [原題]

IN Tachiya, Naohisa
 PA Cosmo Oil Co., Ltd., Japan
 SO Jpn. Kokai Tokkyo Koho, 13pp.
 CODEN: JKXXAF
 DT Patent

STN Tokyo

4/6

LA Japanese

FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI JP 2007238577	A	20070920	JP 2006-66967	20060313
AU 2007237792	A1	20071025	AU 2007-237792	20070301
CA 2618659	A1	20071025	CA 2007-2618659	20070301
WO 2007119302	A1	20071025	WO 2007-JP53962	20070301
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, KE, KG, KM, KN, KP, KR, KZ, LA, LC, JK, LR, LS, LT, LU, LY, MA, MD, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NZ, NO, NZ, OM, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, SV, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW				
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EP 2006279	A1	20081224	EP 2007-737632	20070301
R: AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IS, IT, LI, LT, LU, LV, MC, MT, NL, PL, PT, RO, SE, SI, SK, TR				
IN 2008DN01222	A	20080627	IN 2008-DN1222	20080212
NO 2008000767	A	20080103	NO 2008-767	20080212
KR 2008099229	A	20081112	KR 2008-704709	20080227
CN 101346347	A	20090114	CN 2007-80000942	20080307
PRAI JP 2006-66967	A	20060313		
WO 2007-JP53962	W	20070301		
OS CASREACT 147:386245; MARPAT 147:386245				

L2 ANSWER 3 OF 4 CA COPYRIGHT 2009 ACS on STN

Full Text

AN 146:163393 CA

TI Preparation of 5-aminolevulinic acid ester phosphates and their use for photodynamic therapy and diagnosis, and plant activation

TIJP 5-アミノレブリン酸エステルリン酸塩、その製造方法及びその用途 [原題]

IN Tachiya, Naohisa

PA Cosmo Oil Co., Ltd., Japan

SO Jpn. Kokai Tokkyo Koho, 14pp.

CODEN: JKXXAF

DT Patent

LA Japanese

FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI JP 2007015937	A	20070125	JP 2005-195941	20050705
PRAI JP 2005-195941		20050705		
OS MARPAT 146:163393				

L2 ANSWER 4 OF 4 CA COPYRIGHT 2009 ACS on STN

Full Text

AN 143:422633 CA

TI Preparation of 5-aminolevulinic acid salt, process for producing the same and use thereof

TIJP 5-アミノレブリン酸塩の調製、同じものを生産するためのプロセス、

およびその使用 [機械翻訳]

IN Tachiya, Naohisa; Nishikawa, Seiji; Higo, Mai; Tanaka, Toru; Ishizuka,

STN Tokyo

5/5

Masahiro; Okada, Hideki
 PA Cosmo Oil Co., Ltd., Japan
 SO PCT Int. Appl., 49 pp.
 CODEN: PIIXD2
 DT Patent
 LA Japanese
 FAN.CNT 1

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JP	2006182753	A	20060713	JP 2005-51217	20050225
AU	2005232995	A1	20051027	AU 2005-232995	20050328
CA	2562170	A1	20051027	CA 2005-2562170	20050328
EP	1731500	A1	20061213	EP 2005-727585	20050328
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	JP 2004-99672	A	20040330		
	JP 2004-345661	A	20041130		
	JP 2005-51216	A	20050225		
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	JP 2005-51218	A	20050225		
	WO 2005-JP5765	W	20050328		

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